

Exome Sequence Data From Multigenerational Families Implicate AMPA Receptor Trafficking in Neurocognitive Impairment and Schizophrenia Risk

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Schizophrenia is a mental disorder characterized by impairments in behavior, thought, and neurocognitive performance. We searched for susceptibility loci at a quantitative trait locus (QTL) previously reported for abstraction and mental flexibility (ABF), a cognitive function often compromised in schizophrenia patients and their unaffected relatives. Exome sequences were determined for 134 samples in 8 European American families from the original linkage study, including 25 individuals with schizophrenia or schizoaffective disorder. At chromosome 5q32–35.3, we analyzed 407 protein-altering variants for association with ABF and schizophrenia status. For replication, significant, Bonferroni-corrected findings were tested against cognitive traits in Mexican American families ($n = 959$), as well as interrogated for schizophrenia risk using GWAS results from the Psychiatric Genomics Consortium (PGC). From the gene *SYNPO*, rs6579797 (MAF = 0.032) shows significant associations with ABF ($P = .015$) and schizophrenia ($P = .040$), as well as jointly ($P = .0027$). In the Mexican American pedigrees, rs6579797 exhibits significant associations with IQ ($P = .011$), indicating more global effects on neurocognition. From the PGC results, other *SYNPO* variants were identified with near significant effects on schizophrenia risk, with a local linkage disequilibrium block displaying signatures of positive selection. A second missense variant within the QTL, rs17551608 (MAF = 0.19) in the gene *WWCI*, also displays a significant effect on schizophrenia in our exome sequences ($P = .038$). Remarkably, the protein products of *SYNPO* and *WWCI* are interaction partners involved in AMPA receptor trafficking, a brain process implicated in synaptic plasticity. Our study reveals variants in these genes with significant

effects on neurocognition and schizophrenia risk, identifying a potential pathogenic mechanism for schizophrenia spectrum disorders.

Key words: schizophrenia/cognition/*SYNPO*/*WWCI*/synaptic plasticity

Introduction

Schizophrenia is a complex, highly heritable brain disorder characterized by disturbances in behavior, thought, and emotion.¹ Although a number of genes and neurobiological pathways have been implicated in linkage and genome-wide association studies, much of the genetic liability of schizophrenia remains to be explained,^{2,3} suggesting a polygenic architecture,⁴ likely confounded by the clinical heterogeneity of the disorder.⁵

Deficits in cognitive functions have been commonly observed in schizophrenia patients, and in smaller magnitude in unaffected family members, especially for executive function, learning, and memory,^{6–10} which may reflect innate, underlying differences that mediate the familial risk of schizophrenia. Analysis of such endophenotypes can delineate the psychiatric phenome and allow for identification of etiological mechanisms that are more proximate to gene action than disease endpoints.¹¹ Thus, genes that moderately influence the risk of schizophrenia may exhibit substantially stronger effects on cognition, making them easier to detect at genome-wide significance levels.

This approach was employed by Almasy et al¹² who conducted a genome-wide linkage screen of schizophrenia and cognitive performance in affected families,

discovering a quantitative trait locus (QTL) for abstraction and mental flexibility (ABF) on chromosome 5q (log of odds [LOD] = 3.43; $P = .011$), with effects on schizophrenia risk revealed through bivariate analysis. Although several linkage studies of schizophrenia have implicated chromosome 5q,^{13–20} these findings provide insight into the potential neuropathology of the region, as the neural underpinnings for ABF have been localized primarily to the prefrontal brain circuitry,²¹ including the dorsolateral and superior prefrontal cortices,²² whose anatomical abnormalities and activity levels have been strongly associated with schizophrenia.^{23–27}

The specific genetic variants contributing to this broad linkage region have yet to be determined. Previous endophenotype studies that have targeted schizophrenia candidate genes have been successful in identifying risk variants, including ones for cognitive traits.^{28–38} In this article, we investigate the QTL at chromosome 5q, focusing on nonsynonymous variation from 238 local genes. Exome sequencing was conducted on 134 samples from 8 European American families drawn from the original linkage study, including 25 diagnosed with schizophrenia or schizoaffective disorder. Significant SNP associations for ABF and schizophrenia were followed up in independent Mexican American families for select neurocognitive measurements, as well as examined among the GWAS results from the Psychiatric Genomics Consortium (PGC). We identify a number of potential risk loci, with implications for the neurobiological basis of cognitive impairment as observed in schizophrenia patients.

Methods

Family Samples

The Multiplex-Multigenerational Genetic Investigation of Schizophrenia (MGI) has been described previously.^{12,39} Families were recruited through a European American individual with schizophrenia, who had at least 1 first-degree relative with schizophrenia or schizoaffective disorder (SAD), depressed type. From the extended families, all the available first-, second- and third-degree relatives 15 years of age or older were invited to participate. MGI was approved by the Institutional Review Board of each of the 3 collaborating institutions, with all participants providing informed consent. In the case of minors under age 18 who provided assent, consent was obtained from a parent. Of the 43 MGI families ($n = 676$ participants), 8 of the largest, most densely affected ones were analyzed in this study. A total of 134 samples were exome sequenced, including 23 diagnosed with schizophrenia and 2 with SAD (see [supplementary table S1](#) for pairwise familial relationships). Based on the original ABF linkage,¹² 5 of the families selected exhibit appreciable pedigree-specific LOD scores at the QTL, ranging from 0.16 to 0.33, representing 37.8% of the overall signal (LOD = 3.43; [supplementary table S2](#)).

Phenotyping

DSM-IV diagnoses were determined from: (1) the Diagnostic Interview for Genetics Studies, version 2.0⁴⁰; (2) the Family Interview for Genetics Studies⁴¹; and (3) reviews of medical records. Lifetime best-estimate diagnoses were arrived at by 2 investigators, each blind to the familial relationships among participants ($\kappa > 0.8$). In total, 106 individuals were diagnosed with schizophrenia or SAD, with 75% undergoing treatment at the time of assessment. Effects of medication on neurocognitive measures have been shown to be negligible or subtle.^{42–44} In addition to schizophrenia, other psychiatric conditions identified included schizotypal personality disorder, psychotic disorder, and different forms of bipolar disorder.

Participants completed a computerized test battery^{45,46} designed to evaluate 9 neurocognitive domains.³⁹ ABF was assessed using the Penn Conditional Exclusion Test (PCET),⁴⁷ for which participants are required to select one of 4 shapes for exclusion based on a sorting principle. An efficiency score was calculated as the average z score for performance accuracy and speed.

Exome Sequencing

We used the Illumina TruSeq platform (Illumina) for sample preparation, exome enrichment, and sequencing on the Illumina HiSeq 2000 instrument. In total, 62 Mb were sequenced, yielding uniform coverage of 201 121 exons from 20 794 genes. FASTQ files of demultiplexed paired sequencing reads of 100 bp were produced by CASAVA 1.8 suite and mapped to the UCSC human genome reference assembly 19 (hg19) using BWA (v. 0.6.1).⁴⁸ Mapped reads were analyzed with SAMtools (v. 0.1.12a)⁴⁹ and Picard (v. 1.56) (<http://picard.sourceforge.net>) to mark likely PCR duplicates and ensure consistency of the mapped data, with the output processed with the GATK (v. 1.6) package⁵⁰ (for more detail, see [supplementary methods](#)).

We called a total of 380 895 high-quality SNPs, with an average of 35 reads per variant, each functionally annotated with ANNOVAR.⁵¹ At the QTL for ABF efficiency at 5q32–35.3, spanning approximately 35 Mb, 6518 SNPs were called, encompassing 366 different genes. To focus on sites of potential functional relevance, sequence data were filtered for variant quality LOD scores of 4.0 or greater, have no missing genotype data, and represent nonsynonymous mutations, leaving 407 SNPs available for association analysis. Mendelian consistency of these loci was confirmed with Sequential Oligogenic Linkage Analysis Routines (SOLAR).⁵²

Replication Samples

For any significant, Bonferroni-corrected associations for ABF and/or schizophrenia risk, replications were sought in independent Mexican American families from the Genetics of Brain Structure and Function (GOBSF)

study for select neurocognitive measurements^{53,54}: PCET accuracy ($n = 519$ subjects), Wechsler Adult Intelligence Scale II (WAIS-II; $n = 430$), Wechsler Test of Adult Reading (WTAR) ($n = 264$), California Verbal Learning Test (CVLT) total recall ($n = 520$), and CVLT delayed recall ($n = 518$). Unlike MGI, these families were not ascertained based on schizophrenia probands. Genotypes were obtained from whole genome sequences and imputed data ($n = 959$).⁵⁵

For potential risk effects related to schizophrenia, we interrogated GWAS results from the PGC (available at <http://www.med.unc.edu/pgc/downloads>). Specifically, we examined 2 data sets: stage I, representing 17 population samples of European ancestry ($n = 9394$ cases and 12462 controls), with imputation based on HapMap3 reference panel⁵⁶; and stage I plus additional Swedish cohorts ($n = 5001$ cases and 6243 controls), using 1000 Genomes phase 1 data for imputation.⁵⁷

Statistical Analysis

All genetic analyses were performed in SOLAR, using a maximum likelihood (ML), variance decomposition approach. To evaluate ABF as an endophenotype to schizophrenia, both the genetic correlation and endophenotype ranking variable (ERV) were computed.⁵⁸ SNP association testing was performed using measured genotype analyses.⁵⁹ This single degree of freedom test assumes genetic additivity and compares a model saturated for both the random effects of kinship and the main effect of a SNP genotype to a null model with the SNP effect constrained to 0. Covariates include sex, age, age squared, and their interactions. P values were adjusted for multiple testing ($n = 407$ SNPs) via Bonferroni correction, corresponding to an alpha threshold of approximately 1.2×10^{-4} . Bivariate models of ABF and schizophrenia were also examined for any SNPs of interest. Tail area-based false discovery rate (FDR) q -values were computed in the R package *fdrtool*.⁶⁰ Multimarker, gene-based analyses were conducted using the sequence kernel association tests (SKAT) with the R script *famSKAT*.⁶¹ Diversity and neutrality test statistics were computed for genomic regions of interest using *PopGenome*,⁶² with coalescent simulations (1000 iterations) based on Hudson's MS algorithm⁶³ performed to evaluate significance of observed deviations from the neutral evolutionary model.

Results

Descriptive Statistics and Heritability Estimates

Measures of ABF efficiency were available for 113 of the 134 sequenced individuals, with a mean of -0.45 ± 0.11 , ranging from -2.03 to 1.39 , with no evidence of kurtosis ($g_2 = -1.55$). No significant differences are observed between the sexes. Age is negatively correlated with ABF ($r = -.34$; $P = 2.0 \times 10^{-4}$). For schizophrenia, affected

individuals ($n = 21$) scored significantly worse for ABF efficiency ($\mu = -1.41 \pm 0.24$) than unaffected individuals ($\mu = -0.24 \pm 0.12$; $P = 1.3 \times 10^{-4}$). Both ABF and schizophrenia are significantly heritable, with respective estimates of 0.53 ± 0.19 ($P = 1.8 \times 10^{-3}$) and 0.84 ± 0.40 ($P = 8.6 \times 10^{-3}$). The genetic correlation between the traits is -0.19 ± 0.11 ($P = .34$; ERV = 0.13), with a more robust genetic correlation of -0.47 ± 0.15 ($P = .021$; ERV = 0.28) observed for the entire set of MGI families.

Genetic Associations at 5q32–35.3

In total, 407 nonsynonymous variants from the QTL at 5q32–35.3 were tested for association with ABF ($\lambda = 1.09$; [supplementary table S3](#)). After Bonferroni adjustment for multiple testing, 1 SNP, rs6579797 (MAF = 0.032) from the gene *SYNPO*, was significantly associated with ABF, with the minor allele showing poorer performance ($\beta_{\text{ABF}} = -2.01 \pm 0.48$; $P = 3.70 \times 10^{-5}$; corrected $P = .015$; $q = 0.014$). This SNP, along with 23 others that exhibit nominal association with ABF (ie, $P < .05$), were also tested for association with schizophrenia, of which 7 show evidence of risk ([table 1](#) and [figure 1A](#)). Of these, 2 SNPs remained significant after Bonferroni correction: the top hit for ABF, rs6579797 ($\beta_{\text{SCZ}} = 1.84 \pm 0.63$; $P = .0017$; corrected $P = .040$) and the SNP rs17551608 (MAF = 0.19), located in *WWCI*, with its minor allele associated with improved ABF performance ($\beta_{\text{ABF}} = 0.42 \pm 0.21$; $P = .041$) and decreased liability for schizophrenia ($\beta = -1.09 \pm 0.39$; $P = .0016$; corrected $P = .038$). When considered jointly in a bivariate model, the 2 traits are significantly associated with both rs6579797 ($\beta_{\text{ABF}} = -1.95 \pm 0.47$; $\beta_{\text{SCZ}} = 1.75 \pm 0.55$; $P = 1.11 \times 10^{-4}$) and rs17551608 ($\beta_{\text{ABF}} = 0.45 \pm 0.19$; $\beta_{\text{SCZ}} = -1.00 \pm 0.22$; $P = .0048$). These 2 SNPs are in linkage equilibrium ($r^2 = .019$).

Based on SIFT⁶⁴ and PolyPhen2⁶⁵ algorithms, which predict the effects of amino acid substitutions on protein function, 2 SNPs from [table 1](#) are considered potentially deleterious: rs17551608, and rs17660042 from the gene *SLC36A3* ($P_{\text{ABF}} = .0018$; $P_{\text{SCZ}} = .010$). Interestingly, in addition to rs17551608, 7 other missense variants were identified in *WWCI* ([figure 1B](#)), representing a high concentration of protein-altering variation (top 10th percentile for genes in the region, accounting size). Of these 7 variants, 4 are observed among the 25 schizophrenia cases, 3 of which are predicted to impact protein function. The most noteworthy of these are: rs145963282 (MAF = 0.0083), which is nominally associated with ABF ($\beta_{\text{ABF}} = -2.30 \pm 0.72$; $P = .0029$), and nearly so with schizophrenia risk ($\beta_{\text{SCZ}} = 1.44 \pm 0.82$; $P = .070$; bivariate association $P = .026$) and rs3822659 (MAF = 0.076), which shows increased risk of schizophrenia ($\beta_{\text{SCZ}} = 0.99 \pm 0.41$; $P = .012$). Interestingly, genetic interactions between rs17551608 and these other *WWCI* missense variants were detected, notably rs3822659 for both ABF ($P = .054$)

Table 1. Top Association Results for ABF and Schizophrenia at 5q32–35.3

dbSNP 137	Position (bp)	Gene	MAF	Function Prediction		ABF		Schizophrenia	
				SIFT	PolyPhen2	Beta (SE)	P Value	Beta (SE)	P Value
rs6579797 ^a	149998128	<i>SYNPO</i>	0.032	Tolerated	Benign	-2.01 (0.48)	3.7×10^{-5c}	1.84 (0.63)	.0017 ^c
rs17660042	150666946	<i>SLC36A3</i>	0.073	Deleterious	Probably damaging	-1.02 (0.32)	.0018	0.94 (1.00)	.010
rs2303063	147480027	<i>SPINK5</i>	0.47	Tolerated	Benign	0.41 (0.18)	.023	-0.52 (1.03)	.033
rs2303067	147480955	<i>SPINK5</i>	0.47	Tolerated	Benign	0.41 (0.18)	.023	-0.52 (1.03)	.033
rs2961944	159835658	<i>SLU7</i>	0.20	Tolerated	Benign	-0.40 (0.18)	.027	0.49 (0.24)	.037
rs61740602	150646888	<i>GM2A</i>	0.12	Tolerated	Benign	-0.56 (0.26)	.028	0.79 (0.29)	.0068
rs17551608 ^b	167835539	<i>WWCI</i>	0.19	Deleterious	Possibly damaging	0.42 (0.21)	.041	-1.09 (0.39)	.0016 ^c

Note: The table lists nonsynonymous variants from 5q32–35.3 with at least nominal evidence ($P < .05$) for association with ABF and schizophrenia. SNP rs numbers are based on dbSNP build 137. MAFs are based on maximum likelihood estimates that account for familial relationships. Predicted effects of amino acid changes on protein function are based on the SIFT and PolyPhen2 algorithms, which were obtained with the Ensembl online tool Variant Effect Predictor (VEP). For the association results, positive beta estimates (SE in parentheses) for schizophrenia correspond to increased risk. All 7 of the SNP variants presented here show the expected directions of effect for the 2 traits (ie, decrease in ABF performance corresponds with an increase in schizophrenia risk, and vice versa). ABF, Abstraction and Mental Flexibility; MAFs, Minor Allele Frequencies;

^aG199A; aspartic acid substituted for asparagine, D67N.

^bC798T; arginine substituted for cysteine, R250C.

^cSignificant after Bonferroni correction for multiple testing ($\alpha = .05$): 407 tests for ABF; 24 tests for risk of schizophrenia.

and schizophrenia ($P = .057$), as well as rs61730019 ($P_{ABF} = .039$; $P_{SCZ} = .0042$; [supplementary table S4](#)), with no significant ablation of the main effects of rs17551608.

To assess the independence of these *WWCI* variants, haplotypes were phased using MERLIN v. 1.1.2.⁶⁶ With the exception of rs3822659, which is in perfect linkage disequilibrium (LD) with an adjacent SNP, rs3822660, each of the missense variants were phased to separate haplotypes. Collectively, the haplotypes account for 10.2% of the variation in ABF ($P = 5.95 \times 10^{-5}$) and 20.3% of the risk for schizophrenia (Kullback-Leibler R^2 value; $P = 4.40 \times 10^{-7}$) in our families. Based on multimarker SKAT analyses of ABF, *WWCI* yielded the fifth strongest association among genes tested from the QTL region ($P = .11$; [supplementary table S5](#)), with the lone, nominally significant result belonging to the gene *SLC36A3* ($P = .029$).

Replication of Neurocognitive Effects in GOBSF Families

The 2 SNPs showing significant effects on ABF and/or schizophrenia risk in our MGI families, rs6579797 (*SYNPO*) and rs17551608 (*WWCI*), were tested against neurocognitive measurements obtained in Mexican American pedigrees from GOBSF. Although PCET efficiency, representing a z score of performance accuracy and speed, was not assessed in these independent samples, accuracy scores were available ($n = 519$). However, we found no evidence of association with either rs6579797 ($\beta = -0.041 \pm 0.18$; $P = .82$) or rs17551608 ($\beta = -0.14 \pm 0.12$; $P = .24$).

Given the broad cognitive impairment typically observed in schizophrenia, which extends beyond executive functions, such as ABF, we examined a pair of IQ measures for general intelligence ([table 2](#)), revealing significant, detrimental effects of rs6579797 (WAIS-II:

$\beta = -6.28 \pm 2.79$, $P = .025$; WTAR: $\beta = -7.54 \pm 2.96$, $P = .011$). Also, with previous research strongly implicating *WWCI* in verbal memory performance (see “Discussion” section), we examined CVLT scores, with the *WWCI* variant rs61730019 showing significant association with both delayed recall ($P = .0031$) and total recall ($P = .0091$).

PGC GWAS Results for Schizophrenia

To investigate the effects of *SYNPO* and *WWCI* on schizophrenia, we interrogated GWAS results from the PGC. For stage I analyses ($n = 21\,856$), as reported by Ripke et al,⁵⁶ rs6579797 (*SYNPO*) was neither directly genotyped nor imputed from HapMap 3 data, however a near significant risk effect was observed for a nearby tagging variant ($r^2 = 1.0$), intronic SNP rs9324647 (OR = 4.90 ± 0.91 ; $P = .080$; MAF = 0.0046). More recently, the PGC sample collection has expanded to include large Swedish cohorts (total $n = 32\,143$), for which Ripke et al⁵⁷ have reported updated association results ([supplementary table S6](#)). Based on 1000 Genomes phase 1 data, rs6579797 was successfully imputed but failed to show evidence of risk effects (OR = 1.04 ± 0.070 ; $P = .61$; MAF = 0.02). For other nearby *SYNPO* variants (± 10 kb from rs6579797), a near significant association was identified for a rare intronic SNP, rs192542133 (OR = 0.79 ± 0.08 ; $P = .0025$; corrected $P = .059$; MAF = 0.015). As for *WWCI* variants, including rs17551608, no evidence was found.

Population Genetics and Signatures of Positive Selection

According to 1000 Genomes data, the putative risk allele of rs6579797 shows marked frequency differences between global populations ([supplementary table S7](#)).

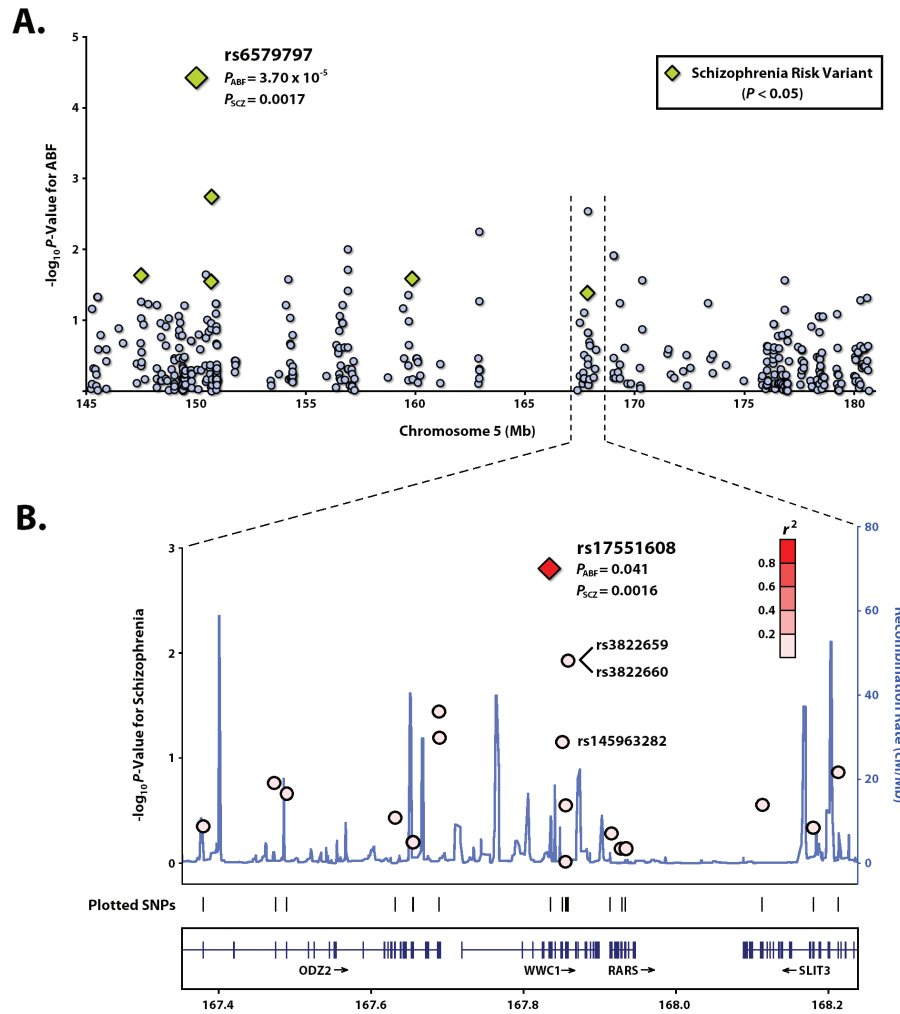


Fig. 1. (A) Plot of association P values for missense SNPs from chromosome 5q32–35.3 ($n = 407$) for ABF. Variants that are also nominally associated with schizophrenia risk ($P < .05$) are represented as diamonds. (B) Regional plot of association results for dbSNP 137 variants from *WWC1* and neighboring genes for schizophrenia. Recombination rate based on hg19 assembly for 1000 Genomes data (2012) for European populations. Plotted using LocusZoom.¹⁴⁹

Table 2. Association Results for rs6579797 and *WWC1* Variants for Select Cognitive Measures in Mexican American Families

			IQ				CVLT			
T2D-GENES Families ($n = 959$)			WASI-II ($n = 430$)		WTAR VIQ ($n = 264$)		Total Recall ($n = 520$)		Delayed Recall ($n = 518$)	
dbSNP 137	Gene	MAF	Beta (SE)	P Value	Beta (SE)	P Value	Beta (SE)	P Value	Beta (SE)	P Value
rs6579797	<i>SYNPO</i>	0.022	-6.28 (2.79)	.025	-7.54 (2.96)	.011	-1.55 (1.96)	.43	-0.38 (0.56)	.50
rs17551608	<i>WWC1</i>	0.077	-2.86 (1.78)	.11	0.74 (1.53)	.63	-1.60 (1.26)	.20	-0.52 (0.36)	.15
rs61730019	<i>WWC1</i>	0.015	1.58 (3.71)	.67	5.24 (3.71)	.16	7.63 (2.57)	.0031	1.93 (0.74)	.0091

Note: The table presents association results between *SYNPO* and *WWC1* variants and measures of IQ and verbal memory in Mexican American families using whole genome sequences and imputed data from the T2D-GENES consortium. Two measures of IQ were tested for genetic association: WASI II and WTAR verbal IQ. For verbal memory, total and delayed recalls for the CVLT were examined. MAFs are based on maximum likelihood estimates that account for familial relationships. Significant association P values ($< .05$) are highlighted in bold. CVLT, California Verbal Learning Test; IQ, Intelligent quotient; MAFs, Minor allele frequencies; WASI, Wechsler Adult Intelligence Scale; WTAR, Wechsler Test of Adult Reading.

Among Europeans (EUR) and admixed Americans (AMR), the respective frequencies are 0.015 and 0.039, whereas higher frequencies are observed for African

(AFR) and Asian (ASN) populations, around 0.25, yielding substantial pairwise F_{ST} scores with the EUR groups (0.22 and 0.21, respectively). As for rs17551608,

population differentiation is also evident, with its minor allele ranging in frequency from 0.16 in EUR populations to its complete absence in the ASN samples.

Interestingly, the minor risk allele (A) of rs6579797 appears to be the ancestral state, as determined from phylogenetic sequence alignments of nonhuman primate species, a possible signature of selection. To investigate this further, we computed the local LD structure of rs6579797 for the 5 EUR populations from 1000 Genomes (figure 2), observing a cluster of *ancestral* minor alleles in strong LD ($r^2 > .9$), indicative of an evolutionary sweep. Diversity and neutrality test statistics support this, as the EUR groups exhibit low nucleotide diversity within the LD block ($\pi = 7.36$), with highly significant deviations from the neutral evolutionary model, including Tajima's D ($P = .010$) and Fu and Li's F ($P = .001$) (table 3 and supplementary table S8). Similar negative deviations from neutrality are observed in admixed American populations, a likely reflection of their European origins.

Discussion

Synaptopodin: Neurocognitive Implications for Schizophrenia Risk

From our analysis of exome sequence data for chromosome 5q32–35.3, a region encompassing a QTL for

neurocognition,¹² we identified nonsynonymous variants from 2 genes, *SYNPO* and *WWCI*, which are significantly associated with ABF and/or schizophrenia risk. Of these, rs6579797 (*SYNPO*) is particularly compelling. The minor allele is carried by 6 heterozygotes in 3 MGI families, representing a modest enrichment relative to EUR populations. Four of the carriers are affected, corresponding to significantly decreased ABF performance (corrected $P = .015$) and heightened risk for schizophrenia (corrected $P = .040$). Based on the original linkage results for ABF, 2 of the 3 families harboring the rs6579797 risk allele have pedigree-specific LODs of 0.33 and 0.28. In the third family, a near zero LOD score was observed, with 2 heterozygote carriers, both unaffected for schizophrenia, although one diagnosed with severe major depression with symptoms of psychosis.

With genome-wide microsatellite data available for many of the MGI samples,¹² we estimated identity-by-descent sharing for chromosome 5, allowing us to impute ML genotypes (.95 probability threshold) for rs6579797 for an additional 82 individuals in our 8 study families (see supplementary methods). Combining the exome sequence data and imputed genotypes, the association signals at rs6579797 remained significant for both ABF (corrected $P = .0031$) and schizophrenia (corrected $P = .024$; supplementary table S9).

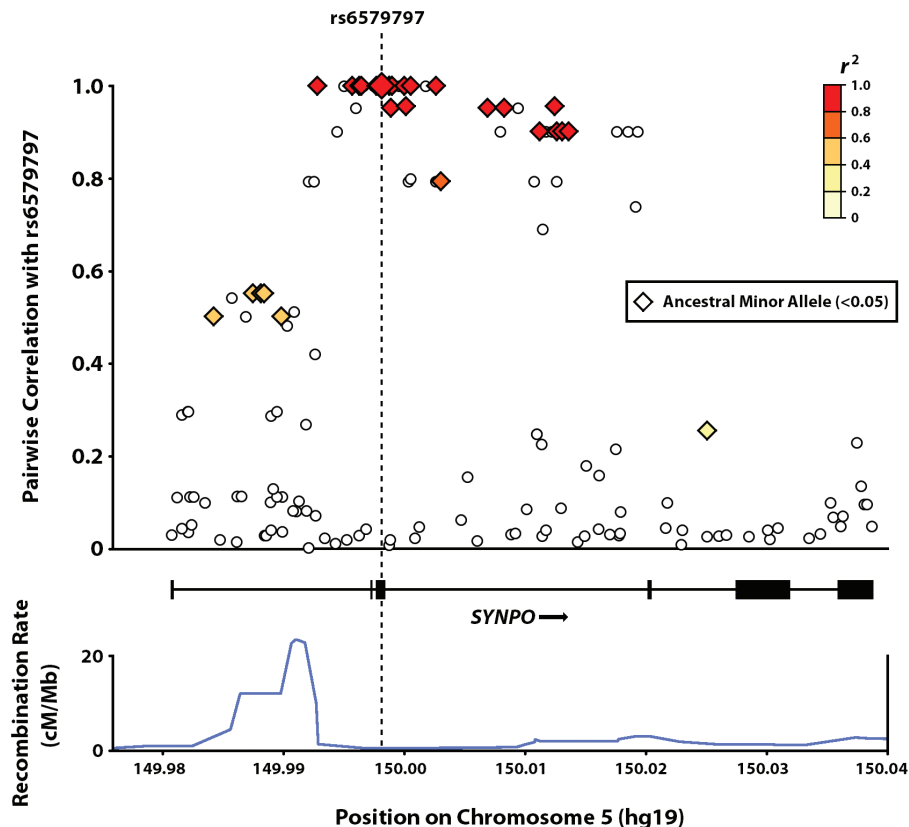


Fig. 2. Plot of pairwise LD correlations for rs6579797 with local *SYNPO* variants, based on population samples of European ancestry from 1000 Genomes data. Ancestral minor alleles (frequency < 0.05) are highlighted.

Table 3. Diversity and Neutrality Test Scores Based on 1000 Genomes Data for LD Block of *SYNPO* Variant rs6579797

1000 Genomes Populations ^a	<i>N</i>	<i>S</i>	π	Tajima's <i>D</i>	Fu and Li's <i>F</i>	Fu and Li's <i>D</i>	Fay and Wu's <i>H</i>	Zeng's <i>E</i>
African	246	211	34.36	-0.04	0.07	0.18	0.73	-0.66
Admixed American	181	156	10.91	-1.89**	-2.83**	-2.83**	0.43 ^c	-2.06**
East Asian	286	107	26.59	1.66	1.53	0.90	-4.44	5.14 ^b
European	379	116	7.36	-1.75**	-3.02**	-3.37**	0.44 ^c	-1.90**
Total	1092	298	23.04	-1.19	-2.15**	-2.81**	0.69	-1.59*

Note: Diversity measurements (segregating sites [*S*], nucleotide diversity [π]), and neutrality test statistics, as determined by differences in unbiased estimators of $\theta = 4 N_e \mu$, were computed for variants found within an observed LD block for rs6579797 in the gene *SYNPO* (pairwise $r^2 > .90$, carrying ancestral minor alleles), corresponding to hg19 coordinates chr5: 149 992 784–150 013 606 (delineated by SNPs rs10074935 and rs61051686).

^a1000 Genomes phase 1 version 3 data for 4 “super populations”: African (AFR), representing Yoruba in Nigeria (YRI), Luhya in Kenya (LWK), and Americans of African ancestry from southwestern United States (ASW); Admixed American (AMR), representing Colombians from Medellin, Colombia (CLM), Puerto Ricans (PUR), and Americans of Mexican ancestry from Los Angeles (MXL); East Asian (ASN), representing Han Chinese from Beijing (CHB), Southern Han Chinese (CHS), and Japanese from Tokyo (JPT); and European (EUR), representing Utah residents with Northern and Western European ancestry (CEU), Toscani in Italy (TSI), British in England and Scotland (GBR), Iberian population in Spain (IBS), and Finnish in Finland (FIN).

^bFor the other tail of the distribution (ie, positive), this score is significant, with an empirical *P* value of .034.

^cPositive deviation Fay and Wu's *H*, a statistic that utilizes information from the intermediate- and high-frequency parts of the frequency spectrum, coupled with the negative score for Zeng's *E* based on low- and high-frequency variant classes, suggests that the locus may be entering a recovery phase for these 2 populations.

*Empirical *P* value (1-tailed) <0.05, ** <0.01, based on an observed distribution for 1000 samples generated via coalescent simulation.

When examined in Mexican American pedigrees from GOBSF, the *SYNPO* variant showed significant association with IQ measures (smallest *P* = .011), indicating more generalized cognitive effects. This is consistent with the overlapping signals observed at the QTL, ranging in LODs from 1.05 to 1.70, for various neurocognitive traits: verbal memory, spatial processing, language and reasoning, and attention. Interestingly, when tested against these other measurements in the MGI families, rs6579797 showed significant detrimental effect on verbal memory accuracy ($\beta = -1.44 \pm 0.45$; *P* = .031). As for schizophrenia risk, we found suggestive evidence for *SYNPO* variants from GWAS results reported by the PGC, although rs6579797 displayed no association in the most current data.

Remarkably, *SYNPO* appears to be under selective pressure, further hinting at its potential relevance. The putative risk allele of rs6579797 represents the ancestral evolutionary state, yet is uncommon in EUR and AMR populations from 1000 Genomes. Pairwise correlations with rs6579797 reveal an LD block enriched with minor ancestral alleles, a potential footprint of an evolutionary sweep. Neutrality test statistics for EUR and AMR support this, revealing significant deviations from the neutral model. Notably, negative deviation was not observed for Fay and Wu's *H*, which, when coupled with the negative score for Zeng's *E*, suggests that the locus may be entering a recovery phase (ie, accumulation of neutral genetic variation).⁶⁷ This finding adds to a growing list of genes implicated in neurocognition and brain development that appear to have undergone selection over the course of human evolution.⁶⁸

Although SIFT and PolyPhen2 yield low probabilities that rs6579797 is damaging, both algorithms have false

negative rates >10%⁶⁹ and thus do not necessarily preclude it from having important functional consequences. The product of *SYNPO*, synaptopodin, is an actin-binding protein found in the dendritic spines of telencephalic neurons,⁷⁰ with the D67N substitution encoded by rs6579797 situated within a PEST motif, which may serve as a molecular signal for proteasomal degradation.⁷¹ Interestingly, when we re-examined *SYNPO* variants excluded from our analyses (eg, those without SIFT and PolyPhen2 scores), we discovered a splice site variant, rs59962087, that is 469 bp upstream of rs6579797 and in perfect LD, with an ancestral minor allele, thus representing another functional candidate for the observed association signal.

What makes this result compelling for the pathology of schizophrenia is that within the dendritic spine, synaptopodin is believed to be an essential component of the spine apparatus (SA), influencing local calcium storage⁷² and protein synthesis,⁷³ with synaptopodin-deficient mice exhibiting deficits in synaptic plasticity and spatial learning.^{74,75} More specifically, synaptopodin directly regulates the release of calcium⁷⁶ and the accumulation of glutamate receptor 1 (GluR1) in the spine head, a subunit of the α -amino-3-hydroxyl-5-methyl-4-isoxazolepropionic acid (AMPA) receptor induced during long-term potentiation (LTP). This establishes a potential mechanistic link with synaptic plasticity,⁷⁷ a key neural process that underlies ABF and mediates synaptic dynamics in the prefrontal cortex (PFC),⁷⁸ a brain region strongly implicated in schizophrenia.^{79–87}

Moreover, in a recent exome sequencing study by Timms et al,⁸⁸ protein-altering variants in genes involved in *N*-methyl-D-aspartate (NMDA) receptor hypofunction

were found to segregate in schizophrenia families, supporting the glutamatergic dysfunction hypothesis for schizophrenia,^{89,90} which ties in well with our findings. NMDA and AMPA are the 2 primary types of receptors activated by glutamate in the mammalian brain, each playing a critical, interrelated role in calcium-induced potentiation. Antagonists of NMDA receptors can replicate schizophrenia symptomatology in healthy people, including deficits in mental flexibility,^{91,92} whereas enhancers have been found to reduce negative features and improve cognition in patients.^{93,94} Studies of knockout mice have revealed impairments in behavioral flexibility,^{95–97} with effects on potentiation.^{98,99} In postmortem brain tissue of schizophrenia patients, altered mRNA and protein levels of glutamate receptors have been observed,¹⁰⁰ including irregularities in AMPA receptor trafficking and localization, particularly in the PFC.^{101–104} Association studies of schizophrenia have identified a number of SNPs and copy number variants (CNVs) in genes involved in glutamatergic neurotransmission,^{105–109} including the synaptic adhesion molecule neurexin,^{110,111} which has been found to regulate AMPA receptor endocytosis and control excitatory synaptic strength.¹¹²

WWCI: Another Regulator of AMPA Receptor Trafficking

The other gene implicated in our analysis, *WWCI*, shows an enrichment of protein-altering variation in our MGI families, including 3 independent SNPs with significant or suggestive associations with ABF and schizophrenia, maintained by the imputed data (supplementary table S9): rs17551608, rs145963282, and rs3822659. Collectively, the *WWCI* variants account for significant portions of the variability in ABF and schizophrenia risk, with evidence of genetic interaction effects. Interestingly, when affection status was broadened to include other schizophrenia spectrum diagnoses in these pedigrees, namely schizotypal personality disorder ($n = 7$) and psychosis disorder ($n = 6$), a stronger association was observed for rs17551608 ($P = 3.80 \times 10^{-4}$; $\beta = -1.07 \pm 0.08$). However, these findings were not independently replicated in the GOBSF families for PCET accuracy and IQ, as well as among the GWAS results for schizophrenia from the PGC, perhaps a reflection of the genetic loads carried by our multiplex families.

Nonetheless, *WWCI* remains intriguing. Its protein product, the WW domain containing protein 1 (WWC1), a postsynaptic scaffolding molecule expressed in the human brain, exhibits protein-protein interactions (PPIs) with other postsynaptic proteins, most notably dendrin and synaptopodin,⁷⁰ via its WW domains.^{113,114} Other binding features include a C2-like motif that interacts with phospholipids¹¹⁵ and a region that binds protein kinase C (PKC) ζ ,¹¹⁶ a molecule integral for neuronal plasticity¹¹⁷ and known to affect long-term memory.¹¹⁸ Based

on the online database BioGRID (v. 3.2.102),¹¹⁹ other PPIs have been detected for synaptopodin and WWC1, including proteins of genes implicated in schizophrenia risk (supplementary table S10).^{120–127} However, no genetic interaction effects were detected between rs6579797 and *WWCI* missense variants for ABF or schizophrenia risk, as well as with other variants from these PPI networks (supplementary table S11).

The influence of *WWCI* on neurocognition, however, is well supported. In a seminal paper by Papassotiropoulos et al,¹²⁸ an intronic SNP within *WWCI*, rs17070145, was reported to be associated with human memory, with allelic differences in hippocampal activations during memory tasks. This finding has been replicated in multiple studies involving both healthy subjects and patients with mild cognitive impairment,^{129–134} with differential effects on memory in psychotic individuals,¹³⁵ as well as a predisposition for late-onset Alzheimer's disease.¹³⁶ Remarkably, rs17070145 has also been linked to cognitive flexibility, with tobacco use possibly modulating this effect,¹³⁷ a notable interaction given its prevalence among schizophrenia patients,¹³⁸ although we found no such effects. However, the chromosome 5 QTL does show suggestive linkage for verbal memory accuracy in MGI, as measured by the Penn Word Memory Test (LOD = 1.50, $P = .0043$), with a near significant association ($\beta = -1.05 \pm 0.55$, $P = .059$) detected for the *WWCI* missense variant rs61730019 (MAF = 0.061), that also displays significant effects on CVLT total recall ($P = .0031$) and delayed recall (.0091) scores in GOBSF.

Despite the convincing case for its neurocognitive implications, how WWC1 controls higher brain function at the molecular level remains to be elucidated. Research has shown that it binds to PICK1,¹³⁹ a synaptic protein involved in AMPA receptor trafficking¹⁴⁰ and considered crucial for hippocampal synaptic plasticity,¹⁴¹ as WWC1 knockdown accelerates the rate of AMPA receptor recycling, with knockout mice exhibiting profound learning and memory impairment. It has been hypothesized that WWC1 may serve as a docking station for AMPA receptors,¹⁴² mediating linkage between endosomes containing phosphatidylinositol-3-phosphate, a key regulator of vesicular traffic in excitatory neurons,¹⁴³ and components of the postsynaptic cytoskeleton that include dendrin and synaptopodin.

Conclusion

From our analysis of exome sequence data from chromosome 5q32–35.3, a region linked to neurocognition in families impacted by schizophrenia, we identify missense variants in 2 genes involved in AMPA receptor trafficking and neuronal plasticity, *SYNPO* and *WWCI*, that are associated with ABF performance and schizophrenia susceptibility. When examined in Mexican American pedigrees, the *SYNPO* variant, rs6579797, shows deleterious effects on

general intelligence, with evidence of selection operating at this locus. Thus, these findings suggest that disruptions in AMPA receptor turnover in the postsynaptic cell have important pathological consequences on neurocognition, lending support to the glutamatergic dysfunction hypothesis. Recognizing that functional improvement in schizophrenia patients is likely to require treatment of cognitive capabilities,^{144,145} as global impairment represents a core feature,¹⁴⁶ augmenting synaptic transmission and plasticity may have therapeutic potential, with a diverse class of allosteric agents available for modulating AMPA receptor activity.¹⁴⁷ Of course, schizophrenia is a highly complex disorder involving perhaps thousands of risk alleles⁴ from genes involved in other neurotransmitter systems, with increasing evidence for the central importance of calcium channel signaling,¹⁴⁸ thus necessitating an integrated, neurobiological approach to developing effective treatment strategies for this devastating mental illness.

Supplementary Material

Supplementary material (references 150–152 are cited in the supplementary material) is available at <http://schizophreniabulletin.oxfordjournals.org>.

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References

1. American Psychiatric Association. *Diagnostic and Statistical Manual of Mental Disorders*. 4th Ed. Washington, DC: American Psychiatric Association; 1994.
2. Visscher PM, Goddard ME, Derks EM, Wray NR. Evidence-based psychiatric genetics, AKA the false dichotomy between common and rare variant hypotheses. *Mol Psychiatry*. 2012;17:474–485.
3. Lee SH, DeCandia TR, Ripke S, et al.; Schizophrenia Psychiatric Genome-Wide Association Study Consortium (PGC-SCZ); International Schizophrenia Consortium (ISC); Molecular Genetics of Schizophrenia Collaboration (MGS). Estimating the proportion of variation in susceptibility

- to schizophrenia captured by common SNPs. *Nat Genet*. 2012;44:247–250.
4. Purcell SM, Wray NR, Stone JL, et al. Common polygenic variation contributes to risk of schizophrenia and bipolar disorder. *Nature*. 2009;460:748–752.
5. Lett TA, Chakravarty MM, Felsky D, et al. The genome-wide supported microRNA-137 variant predicts phenotypic heterogeneity within schizophrenia. *Mol Psychiatry*. 2013;18:443–450.
6. Saykin AJ, Gur RC, Gur RE, et al. Neuropsychological function in schizophrenia. Selective impairment in memory and learning. *Arch Gen Psychiatry*. 1991;48:618–624.
7. Elvevåg B, Goldberg TE. Cognitive impairment in schizophrenia is the core of the disorder. *Crit Rev Neurobiol*. 2000;14:1–21.
8. Egan MF, Goldberg TE, Gscheidle T, et al. Relative risk for cognitive impairments in siblings of patients with schizophrenia. *Biol Psychiatry*. 2001;50:98–107.
9. Conklin HM, Curtis CE, Calkins ME, Iacono WG. Working memory functioning in schizophrenia patients and their first-degree relatives: cognitive functioning shedding light on etiology. *Neuropsychologia*. 2005;43:930–942.
10. Thompson JL, Watson JR, Steinhauer SR, Goldstein G, Pogue-Geile MF. Indicators of genetic liability to schizophrenia: a sibling study of neuropsychological performance. *Schizophr Bull*. 2005;31:85–96.
11. Gottesman II, Gould TD. The endophenotype concept in psychiatry: etymology and strategic intentions. *Am J Psychiatry*. 2003;160:636–645.
12. Almasy L, Gur RC, Haack K, et al. A genome screen for quantitative trait loci influencing schizophrenia and neurocognitive phenotypes. *Am J Psychiatry*. 2008;165:1185–1192.
13. Silverman JM, Greenberg DA, Altstiel LD, et al. Evidence of a locus for schizophrenia and related disorders on the short arm of chromosome 5 in a large pedigree. *Am J Med Genet*. 1996;67:162–171.
14. Straub RE, MacLean CJ, O'Neill FA, Walsh D, Kendler KS. Support for a possible schizophrenia vulnerability locus in region 5q22-31 in Irish families. *Mol Psychiatry*. 1997;2:148–155.
15. Shaw SH, Kelly M, Smith AB, et al. A genome-wide search for schizophrenia susceptibility genes. *Am J Med Genet*. 1998;81:364–376.
16. Gurling HM, Kalsi G, Brynjolfson J, et al. Genomewide genetic linkage analysis confirms the presence of susceptibility loci for schizophrenia, on chromosomes 1q32.2, 5q33.2, and 8p21-22 and provides support for linkage to schizophrenia, on chromosomes 11q23.3-24 and 20q12.1-11.23. *Am J Hum Genet*. 2001;68:661–673.
17. DeLisi LE, Mesen A, Rodriguez C, et al. Genome-wide scan for linkage to schizophrenia in a Spanish-origin cohort from Costa Rica. *Am J Med Genet*. 2002;114:497–508.
18. Devlin B, Bacanu SA, Roeder K, et al. Genome-wide multipoint linkage analyses of multiplex schizophrenia pedigrees from the oceanic nation of Palau. *Mol Psychiatry*. 2002;7:689–694.
19. Sklar P, Pato MT, Kirby A, et al. Genome-wide scan in Portuguese Island families identifies 5q31-5q35 as a susceptibility locus for schizophrenia and psychosis. *Mol Psychiatry*. 2004;9:213–218.
20. Ng MY, Levinson DF, Faraone SV, et al. Meta-analysis of 32 genome-wide linkage studies of schizophrenia. *Mol Psychiatry*. 2009;14:774–785.

21. Krawczyk DC. Contributions of the prefrontal cortex to the neural basis of human decision making. *Neurosci Biobehav Rev.* 2002;26:631–664.
22. Liang P, Wang Z, Yang Y, Jia X, Li K. Functional disconnection and compensation in mild cognitive impairment: evidence from DLPFC connectivity using resting-state fMRI. *PLoS One.* 2011;6:e22153.
23. Weinberger DR, Berman KF, Zec RF. Physiologic dysfunction of dorsolateral prefrontal cortex in schizophrenia. I. Regional cerebral blood flow evidence. *Arch Gen Psychiatry.* 1986;43:114–124.
24. Minzenberg MJ, Laird AR, Thelen S, Carter CS, Glahn DC. Meta-analysis of 41 functional neuroimaging studies of executive function in schizophrenia. *Arch Gen Psychiatry.* 2009;66:811–822.
25. Shashi V, Kwopil TR, Kaczorowski J, et al. Evidence of gray matter reduction and dysfunction in chromosome 22q11.2 deletion syndrome. *Psychiatry Res.* 2010;181:1–8.
26. van Veelen NM, Vink M, Ramsey NF, Kahn RS. Left dorsolateral prefrontal cortex dysfunction in medication-naïve schizophrenia. *Schizophr Res.* 2010;123:22–29.
27. Ursu S, Kring AM, Gard MG, et al. Prefrontal cortical deficits and impaired cognition-emotion interactions in schizophrenia. *Am J Psychiatry.* 2011;168:276–285.
28. Weinshilboum RM, Otterness DM, Szumlanski CL. Methylation pharmacogenetics: catechol O-methyltransferase, thiopurine methyltransferase, and histamine N-methyltransferase. *Annu Rev Pharmacol Toxicol.* 1999;39:19–52.
29. Chen J, Lipska BK, Halim N, et al. Functional analysis of genetic variation in catechol-O-methyltransferase (COMT): effects on mRNA, protein, and enzyme activity in postmortem human brain. *Am J Hum Genet.* 2004;75:807–821.
30. Goldberg TE, Egan MF, Gscheidle T, et al. Executive subprocesses in working memory: relationship to catechol-O-methyltransferase Val158Met genotype and schizophrenia. *Arch Gen Psychiatry.* 2003;60:889–896.
31. Blasi G, Mattay VS, Bertolino A, et al. Effect of catechol-O-methyltransferase val158met genotype on attentional control. *J Neurosci.* 2005;25:5038–5045.
32. Bruder GE, Keilp JG, Xu H, et al. Catechol-O-methyltransferase (COMT) genotypes and working memory: associations with differing cognitive operations. *Biol Psychiatry.* 2005;58:901–907.
33. Aguilera M, Barrantes-Vidal N, Arias B, et al. Putative role of the COMT gene polymorphism (Val158Met) on verbal working memory functioning in a healthy population. *Am J Med Genet B Neuropsychiatr Genet.* 2008;147B:898–902.
34. Burdick KE, Goldberg TE, Funke B, et al. DTNBP1 genotype influences cognitive decline in schizophrenia. *Schizophr Res.* 2007;89:169–172.
35. Baek JH, Kim JS, Ryu S, et al. Association of genetic variations in DTNBP1 with cognitive function in schizophrenia patients and healthy subjects. *Am J Med Genet B Neuropsychiatr Genet.* 2012;159B:841–849.
36. Burdick KE, Hodgkinson CA, Szeszko PR, et al. DISC1 and neurocognitive function in schizophrenia. *Neuroreport.* 2005;16:1399–1402.
37. Carless MA, Glahn DC, Johnson MP, et al. Impact of DISC1 variation on neuroanatomical and neurocognitive phenotypes. *Mol Psychiatry.* 2011;16:1096–1104, 1063.
38. Prasad KM, Almasy L, Gur RC, et al. RGS4 polymorphisms associated with variability of cognitive performance in a family-based schizophrenia sample. *Schizophr Bull.* 2010;36:983–990.
39. Gur RE, Nimgaonkar VL, Almasy L, et al. Neurocognitive endophenotypes in a multiplex multigenerational family study of schizophrenia. *Am J Psychiatry.* 2007;164:813–819.
40. Nurnberger JI, Blehar MC, Kaufmann CA, et al. Diagnostic interview for genetic studies: rationale, unique features, and training. NIMH Genetic Initiative. *Arch Gen Psychiatry.* 1994;51:849–859.
41. Maxwell ME. *Manual for the Family Interview for Genetic Studies (FIGS)*. Bethesda, MD: Clinical Neurogenetics Branch, Intramural Research Program, National Institute of Mental Health; 1992.
42. Saykin AJ, Shtasel DL, Gur RE, et al. Neuropsychological deficits in neuroleptic naïve patients with first-episode schizophrenia. *Arch Gen Psychiatry.* 1994;51:124–131.
43. Censits DM, Ragland JD, Gur RC, Gur RE. Neuropsychological evidence supporting a neurodevelopmental model of schizophrenia: a longitudinal study. *Schizophr Res.* 1997;24:289–298.
44. Goldberg TE, Goldman RS, Burdick KE, et al. Cognitive improvement after treatment with second-generation antipsychotic medications in first-episode schizophrenia: is it a practice effect? *Arch Gen Psychiatry.* 2007;64:1115–1122.
45. Gur RC, Ragland JD, Moberg PJ, et al. Computerized neurocognitive scanning: II. The profile of schizophrenia. *Neuropsychopharmacology.* 2001;25:777–788.
46. Gur RC, Ragland JD, Moberg PJ, et al. Computerized neurocognitive scanning: I. Methodology and validation in healthy people. *Neuropsychopharmacology.* 2001;25:766–776.
47. Kurtz MM, Ragland JD, Moberg PJ, Gur RC. The Penn Conditional Exclusion Test: a new measure of executive-function with alternate forms of repeat administration. *Arch Clin Neuropsychol.* 2004;19:191–201.
48. Li H, Durbin R. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics.* 2009;25:1754–1760.
49. Li H, Handsaker B, Wysoker A, et al.; 1000 Genome Project Data Processing Subgroup. The Sequence Alignment/Map format and SAMtools. *Bioinformatics.* 2009;25:2078–2079.
50. McKenna A, Hanna M, Banks E, et al. The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Res.* 2010;20:1297–1303.
51. Wang K, Li M, Hakonarson H. ANNOVAR: functional annotation of genetic variants from high-throughput sequencing data. *Nucleic Acids Res.* 2010;38:e164.
52. Almasy L, Blangero J. Multipoint quantitative-trait linkage analysis in general pedigrees. *Am J Hum Genet.* 1998;62:1198–1211.
53. Olvera RL, Bearden CE, Velligan DI, et al. Common genetic influences on depression, alcohol, and substance use disorders in Mexican-American families. *Am J Med Genet B Neuropsychiatr Genet.* 2011;156B:561–568.
54. Glahn DC, Kent JW Jr, Sprooten E, et al. Genetic basis of neurocognitive decline and reduced white-matter integrity in normal human brain aging. *Proc Natl Acad Sci USA.* 2013;110:19006–19011.
55. Flannick J, Thorleifsson G, Beer NL, et al.; Go-T2D Consortium; T2D-GENES Consortium. Loss-of-function mutations in SLC30A8 protect against type 2 diabetes. *Nat Genet.* 2014;46:357–363.

56. Ripke S, Sanders AR, Kendler KS, et al. Genome-wide association study identifies five new schizophrenia loci. *Nat Genet.* 2011;43:969–976.
57. Ripke S, O'Dushlaine C, Chambert K, et al.; Multicenter Genetic Studies of Schizophrenia Consortium; Psychosis Endophenotypes International Consortium; Wellcome Trust Case Control Consortium 2. genome-wide association analysis identifies 13 new risk loci for schizophrenia. *Nat Genet.* 2013;45:1150–1159.
58. Glahn DC, Curran JE, Winkler AM, et al. High dimensional endophenotype ranking in the search for major depression risk genes. *Biol Psychiatry.* 2012;71:6–14.
59. Boerwinkle E, Chakraborty R, Sing CF. The use of measured genotype information in the analysis of quantitative phenotypes in man. I. Models and analytical methods. *Ann Hum Genet.* 1986;50:181–194.
60. Strimmer K. fdrtool: a versatile R package for estimating local and tail area-based false discovery rates. *Bioinformatics.* 2008;24:1461–1462.
61. Chen H, Meigs JB, Dupuis J. Sequence kernel association test for quantitative traits in family samples. *Genet Epidemiol.* 2013;37:196–204.
62. Pfeifer B, Wittelsbürger U, Ramos-Onsins SE, Lercher MJ. PopGenome: an efficient Swiss army knife for population genomic analyses in R. *Mol Biol Evol.* 2014;31:1929–1936.
63. Hudson RR. Generating samples under a Wright-Fisher neutral model of genetic variation. *Bioinformatics.* 2002;18:337–338.
64. Kumar P, Henikoff S, Ng PC. Predicting the effects of coding non-synonymous variants on protein function using the SIFT algorithm. *Nat Protoc.* 2009;4:1073–1081.
65. Adzhubei IA, Schmidt S, Peshkin L, et al. A method and server for predicting damaging missense mutations. *Nat Methods.* 2010;7:248–249.
66. Abecasis GR, Cherny SS, Cookson WO, Cardon LR. Merlin—rapid analysis of dense genetic maps using sparse gene flow trees. *Nat Genet.* 2002;30:97–101.
67. Zeng K, Fu YX, Shi S, Wu CI. Statistical tests for detecting positive selection by utilizing high-frequency variants. *Genetics.* 2006;174:1431–1439.
68. Gittelman RM, Hun E, Ay F, et al. Comprehensive identification and analysis of human accelerated regulatory DNA. *Genome Res.* 2015;25:1245–1255.
69. Frousios K, Iliopoulos CS, Schlitt T, Simpson MA. Predicting the functional consequences of non-synonymous DNA sequence variants—evaluation of bioinformatics tools and development of a consensus strategy. *Genomics.* 2013;102:223–228.
70. Mundel P, Heid HW, Mundel TM, Krüger M, Reiser J, Kriz W. Synaptopodin: an actin-associated protein in telencephalic dendrites and renal podocytes. *J Cell Biol.* 1997;139:193–204.
71. Spencer ML, Theodosiou M, Noonan DJ. NPDC-1, a novel regulator of neuronal proliferation, is degraded by the ubiquitin/proteasome system through a PEST degradation motif. *J Biol Chem.* 2004;279:37069–37078.
72. Svoboda K, Mainen ZF. Synaptic $[Ca^{2+}]_i$: intracellular stores spill their guts. *Neuron.* 1999;22:427–430.
73. Pierce JP, van Leyen K, McCarthy JB. Translocation machinery for synthesis of integral membrane and secretory proteins in dendritic spines. *Nat Neurosci.* 2000;3:311–313.
74. Deller T, Korte M, Chabanis S, et al. Synaptopodin-deficient mice lack a spine apparatus and show deficits in synaptic plasticity. *Proc Natl Acad Sci USA.* 2003;100:10494–10499.
75. Jedlicka P, Schwarzscher SW, Winkels R, et al. Impairment of in vivo theta-burst long-term potentiation and network excitability in the dentate gyrus of synaptopodin-deficient mice lacking the spine apparatus and the cisternal organelle. *Hippocampus.* 2009;19:130–140.
76. Korkotian E, Segal M. Synaptopodin regulates release of calcium from stores in dendritic spines of cultured hippocampal neurons. *J Physiol.* 2011;589:5987–5995.
77. Vlachos A, Korkotian E, Schonfeld E, Copanaki E, Deller T, Segal M. Synaptopodin regulates plasticity of dendritic spines in hippocampal neurons. *J Neurosci.* 2009;29:1017–1033.
78. Stokes MG, Kusunoki M, Sigala N, Nili H, Gaffan D, Duncan J. Dynamic coding for cognitive control in prefrontal cortex. *Neuron.* 2013;78:364–375.
79. Goldman-Rakic PS. Working memory dysfunction in schizophrenia. *J Neuropsychiatry Clin Neurosci.* 1994;6:348–357.
80. Eisenberg DP, Berman KF. Executive function, neural circuitry, and genetic mechanisms in schizophrenia. *Neuropsychopharmacology.* 2010;35:258–277.
81. Glantz LA, Lewis DA. Decreased dendritic spine density on prefrontal cortical pyramidal neurons in schizophrenia. *Arch Gen Psychiatry.* 2000;57:65–73.
82. Salisbury DF, Kuroki N, Kasai K, Shenton ME, McCarley RW. Progressive and interrelated functional and structural evidence of post-onset brain reduction in schizophrenia. *Arch Gen Psychiatry.* 2007;64:521–529.
83. Fornito A, Yücel M, Patti J, Wood SJ, Pantelis C. Mapping grey matter reductions in schizophrenia: an anatomical likelihood estimation analysis of voxel-based morphometry studies. *Schizophr Res.* 2009;108:104–113.
84. Zhou Y, Liang M, Jiang T, et al. Functional dysconnectivity of the dorsolateral prefrontal cortex in first-episode schizophrenia using resting-state fMRI. *Neurosci Lett.* 2007;417:297–302.
85. Camchong J, MacDonald AW 3rd, Bell C, Mueller BA, Lim KO. Altered functional and anatomical connectivity in schizophrenia. *Schizophr Bull.* 2011;37:640–650.
86. Cole MW, Anticevic A, Repovs G, Barch D. Variable global dysconnectivity and individual differences in schizophrenia. *Biol Psychiatry.* 2011;70:43–50.
87. Sehatpour P, Dias EC, Butler PD, et al. Impaired visual object processing across an occipital-frontal-hippocampal brain network in schizophrenia: an integrated neuroimaging study. *Arch Gen Psychiatry.* 2010;67:772–782.
88. Timms AE, Dorschner MO, Wechsler J, et al. Support for the N-methyl-D-aspartate receptor hypofunction hypothesis of schizophrenia from exome sequencing in multiplex families. *JAMA Psychiatry.* 2013;70:582–590.
89. Konradi C, Heckers S. Molecular aspects of glutamate dysregulation: implications for schizophrenia and its treatment. *Pharmacol Ther.* 2003;97:153–179.
90. Coyle JT. NMDA receptor and schizophrenia: a brief history. *Schizophr Bull.* 2012;38:920–926.
91. Javitt DC, Zukin SR. Recent advances in the phenylcyclidine model of schizophrenia. *Am J Psychiatry.* 1991;148:1301–1308.
92. Krystal JH, Karper LP, Seibyl JP, et al. Subanesthetic effects of the noncompetitive NMDA antagonist, ketamine, in humans. Psychotomimetic, perceptual, cognitive, and neuroendocrine responses. *Arch Gen Psychiatry.* 1994;51:199–214.
93. Coyle JT, Balu D, Benneyworth M, Basu A, Roseman A. Beyond the dopamine receptor: novel therapeutic

- targets for treating schizophrenia. *Dialogues Clin Neurosci*. 2010;12:359–382.
94. Tsai GE, Lin PY. Strategies to enhance N-methyl-D-aspartate receptor-mediated neurotransmission in schizophrenia, a critical review and meta-analysis. *Curr Pharm Des*. 2010;16:522–537.
 95. Kirchner L, Weitzdoerfer R, Hoeger H, et al. Impaired cognitive performance in neuronal nitric oxide synthase knockout mice is associated with hippocampal protein derangements. *Nitric Oxide*. 2004;11:316–330.
 96. Brigman JL, Feyder M, Saksida LM, Bussey TJ, Mishina M, Holmes A. Impaired discrimination learning in mice lacking the NMDA receptor NR2A subunit. *Learn Mem*. 2008;15:50–54.
 97. Kim JI, Lee HR, Sim SE, et al. PI3K γ is required for NMDA receptor-dependent long-term depression and behavioral flexibility. *Nat Neurosci*. 2011;14:1447–1454.
 98. Brigman JL, Wright T, Talani G, et al. Loss of GluN2B-containing NMDA receptors in CA1 hippocampus and cortex impairs long-term depression, reduces dendritic spine density, and disrupts learning. *J Neurosci*. 2010;30:4590–4600.
 99. Erickson MA, Maramba LA, Lisman J. A single brief burst induces GluR1-dependent associative short-term potentiation: a potential mechanism for short-term memory. *J Cogn Neurosci*. 2010;22:2530–2540.
 100. Weickert CS, Fung SJ, Catts VS, et al. Molecular evidence of N-methyl-D-aspartate receptor hypofunction in schizophrenia. *Mol Psychiatry*. 2013;18:1185–1192.
 101. Toyooka K, Iritani S, Makifuchi T, et al. Selective reduction of a PDZ protein, SAP-97, in the prefrontal cortex of patients with chronic schizophrenia. *J Neurochem*. 2002;83:797–806.
 102. Dracheva S, McGurk SR, Haroutunian V. mRNA expression of AMPA receptors and AMPA receptor binding proteins in the cerebral cortex of elderly schizophrenics. *J Neurosci Res*. 2005;79:868–878.
 103. Beneyto M, Meador-Woodruff JH. Lamina-specific abnormalities of AMPA receptor trafficking and signaling molecule transcripts in the prefrontal cortex in schizophrenia. *Synapse*. 2006;60:585–598.
 104. Hammond JC, McCullumsmith RE, Funk AJ, Haroutunian V, Meador-Woodruff JH. Evidence for abnormal forward trafficking of AMPA receptors in frontal cortex of elderly patients with schizophrenia. *Neuropsychopharmacology*. 2010;35:2110–2119.
 105. Allen NC, Bagade S, McQueen MB, et al. Systematic meta-analyses and field synopsis of genetic association studies in schizophrenia: the SzGene database. *Nat Genet*. 2008;40:827–834.
 106. Bass NJ, Datta SR, McQuillin A, et al. Evidence for the association of the DAOA (G72) gene with schizophrenia and bipolar disorder but not for the association of the DAO gene with schizophrenia. *Behav Brain Funct*. 2009;5:28.
 107. Nicodemus KK, Law AJ, Luna A, et al. A 5' promoter region SNP in NRG1 is associated with schizophrenia risk and type III isoform expression. *Mol Psychiatry*. 2009;14:741–743.
 108. Kirov G, Pocklington AJ, Holmans P, et al. De novo CNV analysis implicates specific abnormalities of postsynaptic signalling complexes in the pathogenesis of schizophrenia. *Mol Psychiatry*. 2012;17:142–153.
 109. Yokley JL, Prasad KM, Chowdari KV, et al. Genetic associations between neuregulin-1 SNPs and neurocognitive function in multigenerational, multiplex schizophrenia families. *Psychiatr Genet*. 2012;22:70–81.
 110. Südhof TC. Neuroligins and neuexins link synaptic function to cognitive disease. *Nature*. 2008;455:903–911.
 111. Reichelt AC, Rodgers RJ, Clapcote SJ. The role of neuroligins in schizophrenia and autistic spectrum disorder. *Neuropharmacology*. 2012;62:1519–1526.
 112. Aoto J, Martinelli DC, Malenka RC, Tabuchi K, Südhof TC. Presynaptic neuroligin-3 alternative splicing trans-synaptically controls postsynaptic AMPA receptor trafficking. *Cell*. 2013;154:75–88.
 113. Duning K, Schurek EM, Schlüter M, et al. KIBRA modulates directional migration of podocytes. *J Am Soc Nephrol*. 2008;19:1891–1903.
 114. Kremerskothen J, Kindler S, Finger I, Veltel S, Barnekow A. Postsynaptic recruitment of Dendrin depends on both dendritic mRNA transport and synaptic anchoring. *J Neurochem*. 2006;96:1659–1666.
 115. Johannsen S, Duning K, Pavenstädt H, Kremerskothen J, Boeckers TM. Temporal-spatial expression and novel biochemical properties of the memory-related protein KIBRA. *Neuroscience*. 2008;155:1165–1173.
 116. Kremerskothen J, Plaas C, Büther K, et al. Characterization of KIBRA, a novel WW domain-containing protein. *Biochem Biophys Res Commun*. 2003;300:862–867.
 117. Sacktor TC. PKMzeta, LTP maintenance, and the dynamic molecular biology of memory storage. *Prog Brain Res*. 2008;169:27–40.
 118. Serrano P, Yao Y, Sacktor TC. Persistent phosphorylation by protein kinase Mzeta maintains late-phase long-term potentiation. *J Neurosci*. 2005;25:1979–1984.
 119. Chatr-Aryamontri A, Breitkreutz BJ, Heinicke S, et al. The BioGRID interaction database: 2013 update. *Nucleic Acids Res*. 2013;41:D816–D823.
 120. Karlsson R, Graae L, Lekman M, et al. MAG11 copy number variation in bipolar affective disorder and schizophrenia. *Biol Psychiatry*. 2012;71:922–930.
 121. Martins-de-Souza D, Gattaz WF, Schmitt A, et al. Prefrontal cortex shotgun proteome analysis reveals altered calcium homeostasis and immune system imbalance in schizophrenia. *Eur Arch Psychiatry Clin Neurosci*. 2009;259:151–163.
 122. Lee SA, Tsao TT, Yang KC, et al. Construction and analysis of the protein-protein interaction networks for schizophrenia, bipolar disorder, and major depression. *BMC Bioinformatics*. 2011;12 Suppl 13:S20.
 123. Jia Y, Yu X, Zhang B, et al. An association study between polymorphisms in three genes of 14-3-3 (tyrosine 3-monooxygenase/tryptophan 5-monooxygenase activation protein) family and paranoid schizophrenia in northern Chinese population. *Eur Psychiatry*. 2004;19:377–379.
 124. Lipska BK, Peters T, Hyde TM, et al. Expression of DISC1 binding partners is reduced in schizophrenia and associated with DISC1 SNPs. *Hum Mol Genet*. 2006;15:1245–1258.
 125. Kang E, Burdick KE, Kim JY, et al. Interaction between FEZ1 and DISC1 in regulation of neuronal development and risk for schizophrenia. *Neuron*. 2011;72:559–571.
 126. Potkin SG, Turner JA, Guffanti G, et al.; FBIRN. A genome-wide association study of schizophrenia using brain activation as a quantitative phenotype. *Schizophr Bull*. 2009;35:96–108.
 127. Wang Q, Charych EI, Pulito VL, et al. The psychiatric disease risk factors DISC1 and TNIK interact to regulate synapse composition and function. *Mol Psychiatry*. 2011;16:1006–1023.

128. Papassotiropoulos A, Stephan DA, Huentelman MJ, et al. Common Kibra alleles are associated with human memory performance. *Science*. 2006;314:475–478.
129. Almeida OP, Schwab SG, Lautenschlager NT, et al. KIBRA genetic polymorphism influences episodic memory in later life, but does not increase the risk of mild cognitive impairment. *J Cell Mol Med*. 2008;12:1672–1676.
130. Nacmias B, Bessi V, Bagnoli S, et al. KIBRA gene variants are associated with episodic memory performance in subjective memory complaints. *Neurosci Lett*. 2008;436:145–147.
131. Schaper K, Kolsch H, Popp J, Wagner M, Jessen F. KIBRA gene variants are associated with episodic memory in healthy elderly. *Neurobiol Aging*. 2008;29:1123–1125.
132. Bates TC, Price JF, Harris SE, et al. Association of KIBRA and memory. *Neurosci Lett*. 2009;458:140–143.
133. Preuschhof C, Heekeren HR, Li SC, Sander T, Lindenberger U, Bäckman L. KIBRA and CLSTN2 polymorphisms exert interactive effects on human episodic memory. *Neuropsychologia*. 2010;48:402–408.
134. Milnik A, Heck A, Vogler C, Heinze HJ, de Quervain DJ, Papassotiropoulos A. Association of KIBRA with episodic and working memory: a meta-analysis. *Am J Med Genet B Neuropsychiatr Genet*. 2012;159B:958–969.
135. Vassos E, Bramon E, Picchioni M, et al. Evidence of association of KIBRA genotype with episodic memory in families of psychotic patients and controls. *J Psychiatr Res*. 2010;44:795–798.
136. Corneveaux JJ, Liang WS, Reiman EM, et al. Evidence for an association between KIBRA and late-onset Alzheimer's disease. *Neurobiol Aging*. 2010;31:901–909.
137. Zhang H, Kranzler HR, Poling J, Gruen JR, Gelernter J. Cognitive flexibility is associated with KIBRA variant and modulated by recent tobacco use. *Neuropsychopharmacology*. 2009;34:2508–2516.
138. Winterer G. Why do patients with schizophrenia smoke? *Curr Opin Psychiatry*. 2010;23:112–119.
139. Makuch L, Volk L, Anggono V, et al. Regulation of AMPA receptor function by the human memory-associated gene KIBRA. *Neuron*. 2011;71:1022–1029.
140. Citri A, Bhattacharyya S, Ma C, et al. Calcium binding to PICK1 is essential for the intracellular retention of AMPA receptors underlying long-term depression. *J Neurosci*. 2010;30:16437–16452.
141. Terashima A, Pelkey KA, Rah JC, et al. An essential role for PICK1 in NMDA receptor-dependent bidirectional synaptic plasticity. *Neuron*. 2008;57:872–882.
142. Duning K, Wennmann DO, Bokemeyer A, et al. Common exonic missense variants in the C2 domain of the human KIBRA protein modify lipid binding and cognitive performance. *Transl Psychiatry*. 2013;3:e272.
143. Shepherd JD, Huganir RL. The cell biology of synaptic plasticity: AMPA receptor trafficking. *Annu Rev Cell Dev Biol*. 2007;23:613–643.
144. Buchanan RW, Freedman R, Javitt DC, Abi-Dargham A, Lieberman JA. Recent advances in the development of novel pharmacological agents for the treatment of cognitive impairments in schizophrenia. *Schizophr Bull*. 2007;33:1120–1130.
145. Barch DM, Carter CS, Arnsten A, et al. Selecting paradigms from cognitive neuroscience for translation into use in clinical trials: proceedings of the third CNTRICS meeting. *Schizophr Bull*. 2009;35:109–114.
146. Schaefer J, Giangrande E, Weinberger DR, Dickinson D. The global cognitive impairment in schizophrenia: consistent over decades and around the world. *Schizophr Res*. 2013;150:42–50.
147. Traynelis SF, Wollmuth LP, McBain CJ, et al. Glutamate receptor ion channels: structure, regulation, and function. *Pharmacol Rev*. 2010;62:405–496.
148. Purcell SM, Moran JL, Fromer M, et al. A polygenic burden of rare disruptive mutations in schizophrenia. *Nature*. 2014;506:185–190.
149. Pruim RJ, Welch RP, Sanna S, et al. LocusZoom: regional visualization of genome-wide association scan results. *Bioinformatics*. 2010;26:2336–2337.